

# ALGOLOGICAL STUDIES OF THE MAROS (MUREŞ) RIVER

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## Introduction

Algae play an important role in riverine ecosystems: they produce oxygen, serve as food for animals, and indicate conditions of and changes to the environment.

At points near the source and upper sections of rivers, attached algae (periphyton) are subsurface as inhabitants of the planktonic environment (pseudoplankton, tychoplankton). Due to both the high velocity of water flow in upper sections of river basins, and high turbidity caused by inorganic particles, diatoms are the main group of algae. A decrease in velocity and/or increasing nutrient load can lead to the dominance of other groups (e.g. green algae) of algae. In the middle and lower sections of rivers real planktonic algae (potamoplankton) can become increasingly dominant.

The first records on algae in the Maros River were provided by Schaarschmidt (1880). Lepşi (1925-26) studied the plankton of the river at Orăştie, and recorded 13 species. Péterfi and Róbert (1958) described two new species of *Cymbella* (*Cymbella subcapitata* and *Cymbella semielliptica*) from samples that were taken at Tîrgu Mureş. Róbert (1960, 1962), specialist in diatomology, studied the diatoms of the closely connected Tîrgu Mureş backwater between 1960-62. In this backwater an interesting mixture of diatoms that are characteristic for different habitats was found: the planktonic *Melosira granulata* var. *angustissima*, the epiphytic *Synedra parasitica* var. *subconstricta* and *Nitzschia sigmoidea*, the alpin-boreal *Pinnularia karelica*, the rheophyl *Ceratoneis arcus* and *Surirella tenera* var. *nervosa* and the halophyl *Cyclotella meneghiniana*, *Epithemia sorex*, *Bacillaria paradoxa* and *Nitzschia hungarica*. He considered most of the species of the identified 92 to be ubiquitous. Róbert (1962) described two new taxa from this backwater (*Pinnularia interrupta* W.Sm. var. *intermedia* Róbert and *Gomphonema augur* Ehr. var. *marisiensis* Róbert). Róbert (1968) studied the diatoms in samples taken from the Maros river at Tîrgu Mureş in 1953.

Diatoms found in the phytobenthos (the term bioderma used in the cited paper) had rheophyl, bentonic, eutrophic and b-mesosaprobic indication values. He characterized the species as having pseudoplanktonic elements; and numerous diatoms had their origin in the saline waters nearby (Singiorgiu de Mureş).

Uherkovich (1971) took samples in 1962 and in 1967 at the mouth of the Maros near Szeged. Rheon-type diatoms dominated, characteristic potamoplankton was not observed even when the water level was low. The total number of individuals ( $1.8-4.8 \times 10^6$  individuum/l) was higher than that in the Tisza River. The presence of *Cyclotella meneghiniana*, *Nitzschia acicularis* and *Nitzschia palea* among the dominating species indicated a high level of pollution. He concluded that although the Mures has a detectable influence on the Tisza, this is not of considerable significance.

Ádámosi et al. (1978) analyzed the algae of the river along a longitudinal section in 1977. Their conclusion was that the phytoplankton which indicate a high level of pollution and hypertrophic conditions have an essential influence on the Tisza River. A significantly increased number of euplanktonic diatoms and green algae was found.

Dobler & Kovács (1981) analyzed the diatoms in the benthos of the Maros River at the mouth. The eu-politrophic indicator *Cyclotella pseudostelligera*, the planktonic *Skeletonema potamos* and *Nitzschia acicularis*, which can be found in polluted waters, were the dominants. The Maros had a considerable influence on the benthic diatom assemblages of the Tisza River.

Vánca (1981) analyzed the other group of algae in parallel samples of the above survey. His conclusion was that the impact of the Maros on the Tisza is the highest among all the tributaries.

Hamar (1991) established that the phytoplankton of the Maros River is characterized by the dominance of  $\mu$ -algae (2-3  $\mu$ ) during the vegetation period. Also, either green algae or *Cyclotella meneghiniana* can be subdominants. When the total number of algae exceeds  $100 \times 10^6$  individuum/l, the water is slightly polluted and politrophic. Impact on the Tisza River is considerable.

### Material and methods

Samples were taken on 15 sampling sites during a longitudinal sampling trip along the Maros in August 1991. Samples were fixed in Lugol's Iodine. Algae were counted under an inverted microscope. An Olympus type microscope was used in identifications.

### Results

Species composition (fig. 1.)

In this study 159 taxa of algae were found in Mures:

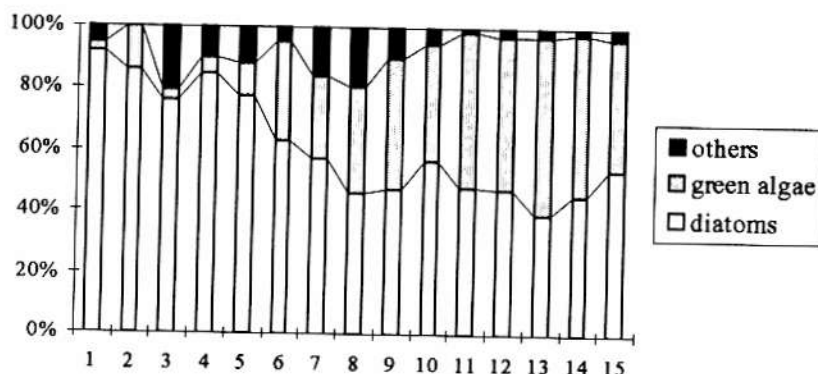
Cyanophyta	10
Euglenophyta	14
Pyrrophyta	7
Chrycophyceae	6
Bacillariophyceae	46
Chlorophyceae	76
Total	159

#### *Cyanophyta*

Blue-green algae are sporadic in upper sections of the river. In the middle and upper sections three species, *Oscillatoria limnetica* Lemm., *Phormidium molle* (Kutz.) Gom. and

*Spirulina laxissima* G.S. West are characteristic. All the three planktonic elements can occur in polluted and saline waters. *Phormidium molle* is saprophytic.

Fig.1. Percentage composition of algal communities



### *Euglenophyta*

Algae belonging to this group are sporadic in the upper and middle sections. Below Alba Iulia species (*Euglena polymorpha* Dang., *Euglena proxima* Dang., *Euglena viridis* Ehr.) indicating polluted conditions appear. The record of *Petalomonas involuta* Skuja is interesting.

### *Pyrrophyta*

They are rare in the upper section, *Rhodomonas lacustris* Pasch. et Rutt. and *Cryptomonas* species are frequently found in lower sections of the river.

### *Chrysophyceae*

Their occurrence is sporadic all along the river basin.

### *Bacillariophyceae*

The upper section is characterized by rheophyl elements, although the number of species that are occurring mostly in streams (like *Achnanthes minutissima* v. *minutissima* Kutz., *Meridion circulare* Ag., *Nitzschia fonticola* Grun.) is rather low. Species indicating eutrophic conditions or moderately polluted environments appear next below the source (*Nitzschia acicularis* (Kutz.) W.Sm., *Navicula cryptocephala* Kutz.). *Nitzschia palea* (Kutz.) W.Sm. is also characteristic in the upper region. Ecological qualification of diatoms lies in a wide range in this river section: oligotrophic and eutrophic, oligosaprobic and a-mesosaprobic indicators are present, they are mostly cosmopolitan. In the midstream section *Cyclotella meneghiniana* Kutz., *Nitzschia palea* (Kutz.) W.Sm., *Nitzschia acicularis* (Kutz.) W.Sm. and *Nitzschia paleacea* Grun. are constant elements. These species indicate eutrophic conditions and polluted environments. Planktonic diatoms appear in this section like *Aulacoseira distans* (Ehr.) Simon., *Acanthoceras zachariaschii* (Brun.) Simon.; and *Nitzschia reversa* W.Sm. which indicate saline waters.

The lower section is similar to the midstream one with more planktonic diatoms like *Aulacoseira granulata* var. *angustissima* (O.F. M.) Simon and *Skeletonema potamos* (Weber) Hasle.

### *Chlorophyta*

Green algae are practically absent in the upper section. A large number of green algae can be found in the middle section, most of them are cosmopolitan and occur in eutrophic waters. *Carteria wisconsinense* H.P. is a rarity, and occurs in planktonic lakes. The number of taxa increases from the upper section, algae belonging to Chlorococcales dominate. A small sized (2-3  $\mu\text{m}$ ) coccoid green alga appears in this region.  $\mu$ -algae invasion has began some years ago in small eutrophic streams, canals and backwaters are found in larger quantities and recently in rivers too. This coccoid green alga regularly occurs in the Maros River in summer (Hamar 1991). Several rare species, like *Chlorogonium elegans* (Dang.) Dang., *Micractinium crassisetum* Hortob., *Polyedropsis spinulosa* (Schmidle) Schmidle and *Pascherina tetras* (Kors.) Silva were also recorded.

### List of algal taxa of the Maros River (1980- )

#### CYANOPHYTA

*Anabaena spiroides* Kleb.  
*Anabaenopsis elenkini* Mull.  
*Aphanizomenon issatschenkoi* (Uss.) Prosch.  
*Microcystis aeruginosa* Kutz.  
*Oscillatoria granulata* Gard.  
*O. limnetica* Lemm.  
*O. prolifica* (Grev.) Gom.  
*Oscillatoria* spp.  
*Phormidium molle* (Kutz.) Gom.  
*Spirulina laxissima* G.S. West

*Gymnodinium excavatum* Nygaard 3.  
*Rhodomonas lacustris* Pasch. et Rutt.

#### CHRYSOPHYTA

CHRYSOPHYCEAE  
*Chrysococcus biporus* Skuja  
*C. rufescens* Klebs  
*Dinobryon divergens* Imhof  
*Mallomonas* sp.  
*Synura globosa* (Schiller) Starmach  
*S. uvella* Ehr.

#### EUGLENOPHYTA

*Euglena acus* Ehr.  
*E. allorgei* Defl.  
*E. geniculata* Duj.  
*E. limnophila* Lemm.  
*E. polymorpha* Dang.  
*E. proxima* Dang.  
*E. viridis* Ehr.  
*Petalomonas involuta* Skuja  
*Phacus arnoldii* Swir.  
*P. pusillus* Lemm.  
*P. pyrum* (Ehr.) Stein 3.  
*P. skujae* Skv.  
*Strombomonas fluviatilis* (Lemm.) Defl.  
*Trachelomonas hispida* (Perty) Stein  
*T. volvocina* Ehr.

#### XANTHOPHYCEAE

*Goniocloris mutica* (A. Braun) Fott 3.

#### BACILLARIOPHYCEAE

*Acanthoceras zachariaschii* (Brun.) Simon.  
*Achnanthes clevei* Grun. 2.  
*A. hungarica* (Grun.) Grun. 2.  
*A. lanceolata* (Bréb.) Grun. 2.  
*A. lanceolata* v. *minor* (Straub) Lange-Bertalot. 2.  
*A. minutissima* v. *minutissima* Kutz.  
*A. plonensis* Hust. 2.  
*Achnanthes* sp.  
*Amphora normanii* Rabh. 2.  
*Amphora spitzbergensis* Van Land. 2.  
*A. pediculus* (Kutz.) Grun.  
*Asterionella formosa* Hass.  
*Aulacoseira distans* (Ehr.) Simon.  
*A. italica* (Ehr.) Simon. 3.  
*A. granulata* (Ehr.) Simon.  
*A. gr.* v. *angustissima* (O.F.M.) Simon.  
*Caloneis amphibaena* (Bory) Cl. 2.  
*Cocconeis neodiminuta* Krammer 2.  
*C. disculus* (Schumann) Cl. 2.

#### PYRROPHYTA

*Cryptomonas curvata* Ehr. em. Pen.  
*C. erosa* Ehr.  
*C. marssonii* Skuja  
*C. obovata* Skuja  
*C. ovata* Ehr.  
*C. reflexa* Skuja

- C. placentula* Ehr.  
*Cyclotella radiosa* (Grun.)Lemm. 2.  
*C. glomerata* Bach. 2.  
*C. meneghiniana* Kutz.  
*C. pseudostelligera* Hust. 2.  
*Cylindrothoea gracilis* (Breb.)Grun. 2.  
*Cymatopleura solea* (Breb.)W.Sm.  
*Cymbella helvetica* Kutz. 2.  
*C. microcephala* Grun. 2.  
*C. silesiaca* Bleisch  
*C. sinuata* Greg. 2.  
*C. silesiaca* Bleisch in Rabenh. 2.  
*Diatoma tenuis* Ag.  
*D. vulgaris* Bory  
*Fragilaria arcus* (Ehr.)Cl. v. *arcus*  
*F. capucina* v. *rumpens*(Kutz.)Lange-Bertalot 2.  
*F. ulna* (Nitzsch)Lange-Bertalot  
*Fragilaria ulna* v. *acus* (Kutz.)Lange-Bertalot  
*Gomphonema angustatum* (Kutz.)Raben.  
*G. augur* Ehr.  
*G. parvulum* (Kutz.)Kutz. 2.  
*G. pseudoaugur* Lange-B.  
*G. olivaceum* (Horn.)Breb.  
*Gyrosigma acuminatum* (Kutz.)Rabh. 2.  
*G. scalproides* (Rabenh.)Cl.  
*M. varians* Ag.  
*Meridion circulare* Ag.  
*N. cari* Ehr. 2.  
*N. cincta* (Ehr.)Ralfs  
*N. cryptocephala* Kutz.  
*N. gregaria* Donk. 2.  
*N. lanceolata* (Ag.)Ehr.  
*N. menisculus* Schuman  
*N. rhynchocephala* Kutz.  
*N. tripunctata* (O.F.M.)Bory 2.  
*N. veneta* Kutz.  
*N. viridula* (Kutz.)Ehr.  
*Nitzschia acicularis* (Kutz.)W.Sm.  
*N. amphibia* Grun. 2.  
*N. constricta* (Kutz.)Ralfs in Pritch.  
*N. dissipata* (Kutz.)Grun.  
*N. fonticola* Grun.  
*N. fruticosa* Hus.  
*N. gracilis* Hantzsch 2.  
*N. hungarica* Grun.  
*N. intermedia* Hantzsch  
*N. palea* (Kutz.)W.Sm.  
*N. paleacea* Grun.  
*N. perminuta* (Grun.)Peral.  
*N. recta* Hantzsch 2.  
*N. reversa* W.Sm.  
*N. subacicularis* Hus.  
*N. sublinearis* Hust. 3.  
*Nitzschia* spp.  
*Rhizosolenia eriensis* H.L.Smith  
*Skeletonema potamos* (Weber)Hasle  
*Stephanodiscus hantzschii* Grun. 2.  
*Stephanodiscus* spp.  
*Surirella angusta* Kutz. 2.
- S. ovalis* Breb.
- CLOROPHYTA**  
*Actinastrum hantzschii* Lagerh.  
*Carteria wisconsinensis* H.P.  
*Chlamydomonas* spp.  
*Chlorogonium elegans* (Dang.)Dang.  
*Closterium acutum* v. *variable* (Lemm.)Krieg. 3.  
*Coccomonas orbicularis* Stein  
*Coelastrum microporum* Naeg.  
*C. sphaericum* Naeg.  
*Cosmarium botrytis* Menegh.  
*Crucigenia apiculata* (Lemm.)Schmidle  
*C. fenestrata* Schmidle  
*C. quadrata* Morr. 3.  
*C. tetrapedia* (Kirschn.)W. et G.S.West  
*Dictyosphaerium anomalum* Kors.  
*D. ehrenbergianum* Naeg. sensu Skuja  
*D. pulchellum* Wood 3.  
*Didymocystis planctonica* Kors.  
*Didymogenes palatina* Schmidle 3.  
*Eudorina elegans* Ehr.  
*Franceia ovalis* (France)Lemm.  
*Golenkinia viridis* (Frenzel)Printz 3.  
*Gonium pectorale* O.F.Muller  
*Granulocytopsis pseudocoronata* (Kors.)Hind. 3.  
*Hyaloraphidium contortum* Pasch. et Kors.  
*H. cont. v. tenuissimum* Kors.  
*Kirchneriella irregularis* ((G.M.)Smith)Kors. 3.  
*Komarekia appendiculata* (Chod.)Fott  
*Korschikoviiella limnetica* (Lemm.)Silva 3.  
*Lagerheimia balatonica* (Scherff.)Hind.  
*L. genevensis* (Chod.)Chod.  
*L. longiseta* (Lemm.)Wille  
*L. quadriseta* (Lemm.)G.M.Smith  
*L. wratislaviensis* Schroed.  
*Lobomonas ampla* v. *mammilata* (Svir.)Kor.  
*Micractinium crassisetum* Hortob.  
*M. pusillum* Fres.  
*Monoraphidium arcuatum* (Kors.)Hind.  
*M. contortum* (Thur.)Kom.-Leg.  
*M. griffithii* (Berk.)Kom.-Leg.  
*M. komarkovae* Nyg.  
*Neodesmus danubialis* Hind.  
*Nephrochlamys willeana* (Printz)Kors.  
*Oocystis borgei* Snow 3.  
*O. lacustris* Chod. 3.  
*O. marssonii* Lemm.  
*Pandorina morum* (O.F.Muller)Bory  
*Pascherina tetras* (Kors.)Silva  
*Pediastrum boryanum* (Turp.)Menegh.  
*P. duplex* Meyen  
*P. simplex* Meyen 3.  
*P. tetras* (Ehr.)Ralfs  
*Polyedropsis spinulosa* (Schm.)Schmidle  
*Scenedesmus acuminatus* (Lagerh.)Chod.  
*S. acutus* Meyen  
*S. apiculatus* (W. et G.S.West)Chod.  
*S. armatus* Chod.

*S. bicaudatus* Dedus.  
*S. brevispina* (G.M.Smith)Chod.  
*S. brevispina* v. *bicaudatus* Hortob.  
*S. denticulatus* Lagerh.  
*S. denticulatus* v. *linearis* Hangs.  
*S. dispar* (Breb.)Rabenh.  
*S. ecornis* (Ehr.)Chod.  
*S. ecornis* v. *disciformis* Chod.  
*S. ellipsoideus* Chod.  
*S. intermedius* Chod.  
*S. intermedius* v. *bicaudatus* Hortob.  
*S. longispina* Chod.  
*S. magnus* Meyen  
*S. opoliensis* P.Richt  
*S. ovalternus* Chod.  
*S. protuberans* Fritsch

*S. quadricauda* (Turp.)Breb.  
*S. spinosus* Chod.  
*Schroederia indica* Phil. 3.  
*S. setigera* (Schroed.)Lemm.  
*S. spiralis* (Printz)Kors.  
*Scourfieldia cordiformis* Takeda  
*Staurostrum paradoxum* Meyen  
*Tetraedron arthrodesmiforme* (West)Wol.  
*T. caudatum* (Corda)Hangs.  
*T. minimum* (A.Br.)Hangs  
*T. proteiforme* (Turn)Brun. 3.  
*T. triangulare* Kors. 3.  
*Tetraselmis cordiformis* (Carter)Stein  
*Tetrastrum glabrum* (Roll)Ahl. et Tiff.  
*T. punctatum* (Schmidle)Ahl. et Tiff.  
*T. staurogeniaeforme* (Schroed.)Lemm.

$\mu$  alga (2-3  $\mu$ ); markless: this study; 2. Dobler-Kovács (1981) only; 3. Hamar (1991) only

### Quantitative changes

In the upper section of the river low numbers of individuals were found (0.06 - 0.14 x 10<sup>6</sup> ind/l) and diatoms dominated (72-92 %). Eutrophic indicator species: *Nitzschia palacea* Grun., *Nitzschia acicularis* (Kutz.) W.Sm. and *Nitzschia palea* (Kutz.) W.Sm. dominated. (Table 1, Fig. 2)

Diatoms remain the dominant group in the middle section; they contribute to total numbers by more than 50% . Dominants: *Cyclotella meneghiniana* Kutz. and the three diatoms listed before. Contribution of green algae is around 30% . Total numbers is higher (0.08 - 0.75 x 10<sup>6</sup> ind/l) than in upper sections. Diatoms and green algae almost equally contribute to total numbers in the lower river sections. *Cyclotella meneghiniana*, *Stephanodiscus* spp., *Nitzschia acicularis* and green  $\mu$ -algae are important. Total numbers changed between 21.5 - 55 x 10<sup>6</sup> ind/l, which indicates that the water is eu-polytrophic and moderately polluted.

### Ecological considerations

Composition of algal communities reflect both the hydrographical properties of the rivers and the effects of allochthonous factors, like pollution (Figs. 2-3). The quickly running (50 - 110 cm/s) Maros receives many small streams in its upper section. Correspondingly, algal abundance is low, diatoms dominate. However, species that indicate pollution appear in this section.

Flow velocity is lower in the middle section (20 - 30 cm/s). Beside rheophyl diatoms, planktonic diatoms and green algae are increasingly dominant. Composition of the algal assemblage indicates considerable pollution. There is a further decrease in velocity (5 - 25 cm/s) in the lower section, in addition the pollution is significant. A large number of

planktonic species that characterize eu-polytrophic conditions and moderate pollution can be experienced (Figs. 1-3).

Fig. 2. Dinamisms of the taxa and number of algae

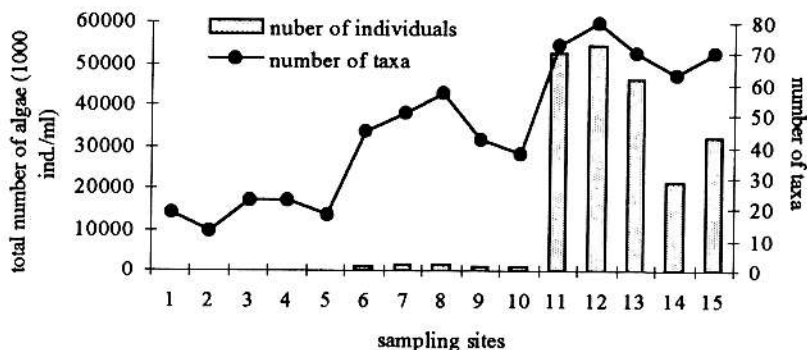
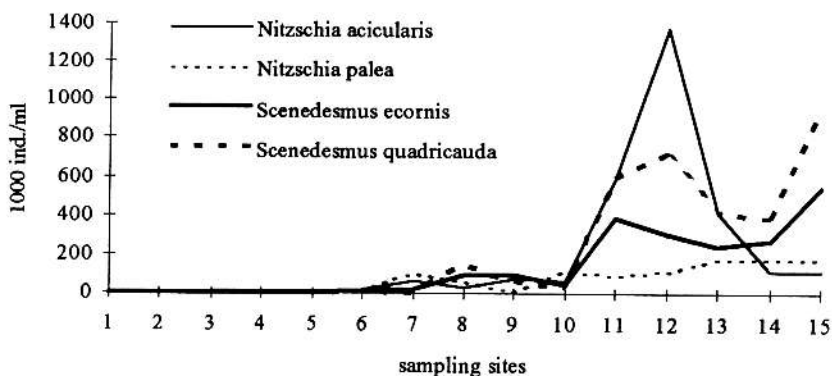


Fig. 3. Indications of pollution by increase of individual number of some algal taxa



The above described phenomena are also reflected in correlation analyses. Algal numbers and number of species positively correlated ( $r = 0.85$ ), while both are negatively correlated with Shannon-diversity ( $r = -0.64$  and  $r = -0.86$ , respectively). The considerable decrease in diversity in the lower section clearly indicates the immense changes in community structure which is caused by pollution. From Alba Iulia - from this point,

Table1. Quantitative dinamism of phytoplankton of the Maros river

	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15
<b>CYANOPHYTA</b>															
<i>Anabaena spiroides</i> Kleb.								3			6				6
<i>Anabaenopsis elenkinii</i> Mull.										6			30		
<i>Aphanizomenon issatschenkoi</i> (Uss.)Pros.							3	6							
<i>Microcystis aeruginosa</i> Kutz.												6			
<i>Oscillatoria granulata</i> Gard.												6			
<i>O. limnetica</i> Lemm.							6	210	75	30	60	6		6	6
<i>O. prolifica</i> (Grev.)Gom.															6
<i>Oscillatoria</i> spp.	1.5		3	3	6			9	3	3					
<i>Phormidium molle</i> (Kutz.)Gom.					1.5						390	1200	960	120	600
<i>Spirulina laxissima</i> G.S.West								60	9	3	30	30			
<b>total Cyanophyta</b>	<b>1.5</b>		<b>3</b>	<b>3</b>	<b>7.5</b>		<b>9</b>	<b>288</b>	<b>87</b>	<b>42</b>	<b>486</b>	<b>1248</b>	<b>990</b>	<b>126</b>	<b>618</b>
<b>EUGLENOPHYTA</b>															
<i>Euglena acus</i> Ehr.						3		3							
<i>E. allorgei</i> Defl.											6				
<i>E. geniculata</i> Duj.						3	3								
<i>E. limnophila</i> Lemm.											6	6	30		6
<i>E. polymorpha</i> Dang.								6		6	30	6	6		6
<i>E. proxima</i> Dang.			1.5				3	3	3	3		30	30	30	6
<i>E. viridis</i> Ehr.											30	30			
<i>Petalomonas involuta</i> Skuja						3									
<i>Phacus arnoldii</i> Swir.	1.5														
<i>P. skujae</i> Skv.							3								
<i>P. pusillus</i> Lemm.							3								
<i>Strombomonas fluviatilis</i> (Lemm.)Defl.														6	
<i>Trachelomonas hispida</i> (Perty)Stein														6	
<i>T. volvocina</i>						3		3	3		30		6		6
<b>total Euglenophyta</b>	<b>1.5</b>			<b>1.5</b>		<b>12</b>	<b>12</b>	<b>15</b>	<b>6</b>	<b>9</b>	<b>102</b>	<b>72</b>	<b>72</b>	<b>42</b>	<b>24</b>



Table 1. (continued)

PYRROPHYTA													
<i>Cryptomonas curvata</i> Ehr. em. Pen.					3	3	3	3	30	24	60	60	60
<i>C. erosa</i> Ehr.									6	6	6	18	6
<i>C. marssonii</i> Skuja	1.5	1.5											
<i>C. obovata</i> Skuja	1.5	3	3		6					90	60	60	180
<i>C. ovata</i> Ehr.	1.5									6	6	120	60
<i>C. reflexa</i> Skuja				3		3							6
<i>Rhodomonas lacustris</i> Pasch. et Rutt.				9	150	3			60	30	6	30	6
<b>total Pyrrophyta</b>	<b>4.5</b>	<b>4.5</b>	<b>3</b>	<b>12</b>	<b>159</b>	<b>9</b>	<b>3</b>	<b>3</b>	<b>96</b>	<b>156</b>	<b>138</b>	<b>288</b>	<b>318</b>
CHRYSOPHYCEAE													
<i>Chrysococcus biporus</i> Skuja				3	3	3							
<i>C. rufescens</i> Klebs	4.5			15	24				30	30			
<i>Dinobryon divergens</i> Imhof	1.5			3									
<i>Mallomonas</i> sp.	9		1.5										
<i>Synura globosa</i> (Schiller)Starmach	4.5												
<i>S. uvella</i> Ehr.				3	3								
<b>total Chrysophyceae</b>	<b>19.5</b>	<b>1.5</b>	<b>24</b>	<b>30</b>	<b>3</b>				<b>30</b>	<b>30</b>			
BACILLARIOPYCEAE													
<i>Acanthoceras zachariaschii</i> (Brun.)Simon.						3					6	6	18
<i>Achnanthes minutissima</i> v. <i>minutissima</i> Kutz.		4.5	1.5	435	300	90	45		150	90	300	6	6
<i>Achnanthes</i> sp.							6						
<i>Amphora pediculus</i> Grun.						6							
<i>Asterionella formosa</i> Hass.													6
<i>Aulacoseira distans</i> (Ehr.)Simon.							135	30		12	120	6	6
<i>A. granulata</i> (Ehr.)Simon.				3	3	9		3	6	12	60	90	60
<i>A. gr. v. angustissima</i> (O.Mull.)Simon.							6		30	6	120	30	90
<i>Cocconeis placentula</i> Ehr.	1.5	4.5	3		6	3							
<i>Cyclotella</i> cf. <i>meneghiniana</i> Kutz.	1.5	3	1.5	150	195	540	105	120	22500	18675	9600	7800	16000
<i>Cymatopleura solea</i> (Breb.)W.Sm.			1.5										
<i>Cymbella silesiaca</i> Bleisch		1.5											
<i>Diatoma tenuis</i> Ag.											12	6	
<i>D. vulgaris</i> Bory								3					
<i>Fragilaria ulna</i> (Nitzsch)Lange-Bertalot	1.5	6			6	3					6		

Table 1. (continued)

<i>F. ulna v. acus</i> (Kutz.) Lange-Bertalot	3										30	60	6		
<i>Gomphonema angustatum</i> (Kutz.) Raben.	1.5		4.5	7.5		3									
<i>G. augur</i> Ehr.	1.5	1.5	1.5												
<i>G. pseudoaugur</i> Lange-Bertalot															
<i>G. olivaceum</i> (Horn.) Breb.														6	
<i>Melosira varians</i> Ag.			1.5		1.5										
<i>Meridion circulare</i> (Grev.) Ag.				1.5											
<i>Navicula cincta</i> (Ehr.) Ralfs								3			6				
<i>N. cryptocephala</i> Kutz.	1.5	1.5	3			3						6			
<i>N. lanceolata</i> (Ag.) Ehr.	1.5														
<i>N. menisculus</i> Schuman														6	
<i>N. rhynchocephala</i> Kutz.	3		3	1.5	1.5	3	3	3	9		60	6			
<i>N. veneta</i> Kutz.		1.5													
<i>N. viridula</i> (Kutz.) Ehr.	3			1.5		3			6						
<i>Nitzschia acicularis</i> (Kutz.) W. Sm.	4.5		7.5	6	9	18	60	30	75	60	600	1380	420	120	
<i>N. dissipata</i> (Kutz.) Grun.		1.5													
<i>N. fonticola</i> Grun.	4.5	9	7.5	1.5						3					
<i>N. fruticosa</i> Hus.											6	30	60	6	
<i>N. hungarica</i> Grun.						3				3				18	
<i>N. intermedia</i> Hantzsch		24								3					
<i>N. palea</i> (Kutz.) W. Sm.		4.5	10.5	4.5	4.5	24	105	60	6	120	90	120	180	180	
<i>N. paleacea</i> Grun.	9	10.5	36	28.5	27	3	27	6	15	105	6	30			
<i>N. perminuta</i> (Grun.) Peral.	6	4.5	7.5	1.5											
<i>N. reversa</i> W. Sm.						6	6	6		30	30	6	6	60	
<i>N. subacicularis</i> Hus.						3	3	3	12	30	30	60	6	60	
<i>Nitzschia</i> spp.	7.5	4.5	6	3	1.5	15	24	3	6	45	60	6	12	60	
<i>Rhizosolenia eriensis</i> H.L. Smith											6	6	6	6	
<i>Skeletonema potamos</i> (Weber) Hasle											18	30	6	6	
<i>Stephanodiscus</i> spp.											1800	5400	7200	1200	
<i>Surirella ovalis</i> Breb.		1.5	1.5	1.5	1.5										
other diatoms	3	1.5	12	1.5	18		3	3	9	6	6	12	6	60	
<b>total Bacillariophyceae</b>	<b>54</b>	<b>66</b>	<b>102</b>	<b>75</b>	<b>76.5</b>	<b>669</b>	<b>750</b>	<b>762</b>	<b>444</b>	<b>558</b>	<b>25338</b>	<b>25971</b>	<b>18192</b>	<b>9726</b>	<b>16322</b>

Table 1. (continued)

## CHLOROPHYTA

Actinastrum hantzschii Lagerh.										6	60	60		
Carteria wisconsinensis H.P.					3	3								
Chlamydomonas spp.	1.5	10.5	4.5	1.5	135	60	60	9	9	120	160	240	120	180
Chlorogonium elegans (Dang.)Dang.										6	30			
Coccomonas orbicularis Stein														
Coelastrum microporum Naeg.						3		6						
C. sphaericum Naeg.							3			12	120	120	12	30
Cosmarium botrytis Menegh.										6	12	60		6
Crucigenia apiculata (Lemm.)Schmidle					3	3	6			6	90			
C. fenestrata Schmidle														
C. tetrapedia (Kirschn.) W. et G.S.West					3	3	6		3				60	6
Dictyosphaerium anomalum Kors.														6
D. ehrenbergianum Naeg. sensu Skuja										30				6
Didymocystis planctonica Kors.										150	150	60	60	
Eudorina elegans Ehr.					6		3	3	6	60	90		12	60
Franceia ovalis (France)Lemm.						3						6		
Gonium pectorale O.F.M.											30			
Hyaloraphidium contortum Pasch. et Kors.					3									
H. c. v.tenuissimum Kors.								9	3					
Komarekia appendiculata (Chod.)Fott					3	3								
Lagerheimia balatonica (Scherff.)Hind.											6	6	90	60
L. genevensis (Chod.)Chod.														
L. longiseta (Lemm.)Wille										12	30	60		
L. quadriseta (Lemm.)G.M.Smith											30			
L. wratislaviensis Schroed.									3					
Lobomonas ampla v.mammilata (Svir.)Kor.										6	60			30
Micractinium crassisetum Hortob.										6	6			
M. pusillum Fres.										30				
										60	60	6	6	30

Table 1. (continued)

<i>Monoraphidium arcuatum</i> (Kors.)Hind.			3	3		30				
<i>M. contortum</i> (Thur.)Kom.-Leg.	24	105	90	3	45	810	540	1140	420	780
<i>M. griffithii</i> (Berk.)Kom.-Leg.	3	3	3	3	6	30	90	160	60	6
<i>M. komarkovae</i> Nyg.	45	24	3	9		30	30	240	60	60
<i>Neodesmus danubialis</i> Hind.	3	3				6				
<i>Nephrochlamys willeana</i> (Printz)Kors.	3		3	3		6		60		30
<i>Oocystis marssonii</i> Lemm.							6		6	60
<i>Pandorina morum</i> (O.F.M.)Bory										6
<i>Pascherina tetras</i> (Kors.)Silva						6	12			
<i>Pediastrum boryanum</i> (Turp.)Menegh.			3	3			30	60	6	
<i>P. duplex</i> Meyen				3			60	60		
<i>P. tetras</i> (Ehr.)Ralfs								6		60
<i>Polyedropsis spinulosa</i> (Schmidle)Schmidle						12			6	30
<i>Scenedesmus acuminatus</i> (Lagerh.)Chod.		3	3	3		90	180	60	6	90
<i>S. acutus</i> Meyen				3		12			6	
<i>S. apiculatus</i> (W. et G.S.West)Chod.								6		
<i>S. armatus</i> Chod.		3								
<i>S. bicaudatus</i> Dedus.		3	6	3	9	30		60	6	30
<i>S. brevispina</i> (G.M.Smith)Chod.			3		3					
<i>S. b. v.bicaudatus</i> Hortob.						6	30		6	30
<i>S. denticulatus</i> Lagerh.			3			6	30	6	30	
<i>S. d. v.linearis</i> Hangs.			3				6	60	6	60
<i>S. dispar</i> (Breb.)Rabenh.					3					
<i>S. eornis</i> (Ehr.)Chod.	9	24	90	90	45	390	300	240	270	540
<i>S. e. v.disciformis</i> Chod.										1.5
<i>S. ellipsoideus</i> Chod.									6	
<i>S. intermedius</i> Chod.		3	3	3		6	120	180	60	120
<i>S. i. v.bicaudatus</i> Hortob.			3			6	6			
<i>S. longispina</i> Chod.				3	3	6	6	6		6
<i>S. magnus</i> Meyen					3	30	6	6	60	30
<i>S. opoliensis</i> P.Richt						60	6	6		120
<i>S. ovalternus</i> Chod.		3					60	60	90	6

Table 1. (continued)

<i>S. protuberans</i> Fritsch								3		9		60	60	12	120	
<i>S. quadricauda</i> (Turp.)Breb.		6	3	150	60	30	600	720	420	390	960					
<i>S. spinosus</i> Chod.		9	9	12	3	6	240	180	240	210	240					
<i>Scourfieldia cordiformis</i> Takeda															60	
<i>Schroederia setigera</i> (Schroed.)Lemm.				3			12	30	60	6	60					
<i>S. spiralis</i> (Printz)Kors.		3		3			6	12	30		6					
<i>Staurastrum paradoxum</i> Meyen		3	3					6								
<i>Tetraedron arthrodesmiforme</i> (West)Wol.							30	60							6	
<i>T. caudatum</i> (Corda)Hangs.								30				30		6		
<i>Tetraedron minimum</i> (A.Br.)Hangs.		6	3	6			30	120						12	6	
<i>Tetraselmis cordiformis</i> (Carter)Stein		3	3	3									6			
<i>Tetrastrum glabrum</i> (Roll)Ahl. et Tiff.		3			3	3	30	90	6	60	60					
<i>T. punctatum</i> (Schmidle)Ahl et Tiff.		3		3			60	120	6							
<i>T. staurogenieforme</i> (Schroed.)Lemm.				3			30		6	6	120					
other Chlorococcales u alga (2-3 u)		1.5	10.5	60	75	90	180	180	240	120						
total Chlorophyta		1.5	10.5	4.5	4.5	10.5	339	351	573	402	369	23250	23400	22630	9000	9000
<b>TOTAL NUMBER OF ALGAE</b>		<b>58.5</b>	<b>76.5</b>	<b>134</b>	<b>88.5</b>	<b>99</b>	<b>1044</b>	<b>1284</b>	<b>1653</b>	<b>942</b>	<b>981</b>	<b>52668</b>	<b>54877</b>	<b>46534</b>	<b>21534</b>	<b>32410</b>
Number of taxa		19	13	23	23	18	45	51	57	42	38	73	80	70	63	70
H' = diversity (Shannon-Weaver)		3.9	3	3.8	3.8	3.3	3.3	3.9	3.7	4	4.1	2	2.4	2.5	2.5	2.8
S = saprobity index (Pantle-Buck)		2.14	2.07	2.05	2	1.83	2.25	2.35	2.2	1.95	2.16	2	2.34	2.27	2.1	2.2

which marks the beginning of the lower section of the river, phytoplankton structures strike one more as a well-operated sewage oxidation pond than a river.

### Conclusions

Composition of algal communities of the Maros River well reflects both the hydro-geological background and human impacts. Surroundings of the heleocean type source has been already slightly polluted. In its upper region the river flows through a basin with only a slight slope (Giurgeu Basin), where the level of pollution increases. In the Toplița-Deda strait self-purification occurs.

The middle section begins above the river dam at Tîrgu Mureș. Downstream impacts of sewage water from Tîrgu Mureș and Luduș can be observed. The two considerable streams, Arieș and Tîrnava, dilute the river.

The lower section begins at Alba Julia, where the river receives a high level of pollution. This leads to an algal community structure that characterizes sewage oxidation ponds. The water quality slightly improves in the lower section, where sewage from Makó contributes to an increase in the level of pollution.

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